

NIPA for Clinically Significant Fetal Antigens

Updated September 2023 Version 3.0

What is NIPA?

Non-Invasive Prenatal Analysis (NIPA) for Clinically Significant Fetal Antigens is a molecular blood group genotyping assay used to predict the blood group phenotype of the fetus. It is performed for cases where the mother is antigen negative and has a corresponding allo-antibody placing the fetus at risk of being affected by Haemolytic Disease of the Fetus and Newborn (HDFN). NIPA utilises maternal peripheral whole blood EDTA samples. Cell-free DNA (cfDNA), which contains cell-free fetal DNA (cffDNA) is extracted from maternal plasma. The DNA is analysed for the presence of the gene or molecular variation responsible for (or associated with) the blood group phenotype under investigation. This technique replaces the current requirement for invasive direct sampling methods for fetal DNA, such as amniocentesis or chorionic villus sampling (CVS) which pose a risk to the pregnancy.

What do we test for?

After extracting cfDNA from the maternal plasma, a droplet digital Polymerase Chain Reaction (ddPCR) assay is used to predict the fetal blood group phenotype for the antigens of interest – D, C, c, E, K, k, Fya, Fyb.

Table 1: ddPCR assays to predict fetal antigen status

Assay name	Gene	Allele/s	Target	Ag prediction	Housekeeper
RHD	RHD	RHD*01/RHD*01N	Exon 5 and Exon 10	D+/D-	CCR5
KEL	KEL	KEL*01.01/KEL*02	rs8176058 578C>G	K+/- or k +/-	KEL*02 or KEL*01.01
FY	ACKR1	FY*01/FY*02	rs12075 125A>G	Fy ^a or Fy ^b	FY*01 or FY*02
RHCE*c	RHCE	RHCE*c	rs676785 307C	C+/-	CCR5
RHCE*E	RHCE	RHCE*E	rs609320 676G>C	E+/-	CCR5
RHCE*C	RHCE	RHCE*C	RHCE intron 2 109bp insert associated with C antigen expression	C+/-	CCR5

How do we test?

The assay is performed using the Bio-Rad Droplet Digital PCR system (ddPCR). This system uses custom designed primer and probe sets for each of the specific targets, which are added together with ddPCR Supermix in a reaction mixture and partitioned into approximately 20,000 individual droplets using the Automated Droplet Generator (AutoDG). Each patient cfDNA sample is tested in five replicates for the fetal antigen of interest, therefore up to 100,000 droplets are potentially generated. Maternal genomic DNA (gDNA) is also tested in a single replicate. Partitioned droplets are then amplified using the C1000 Touch thermal cycler. Amplified partitions are read using the ddPCR QX200 Reader and analysed using QuantaSoft Pro software to determine if fetal antigen signals are present.

Performance

The assay has a positive predictive value (PPV) ≥98% and negative predictive value (NPV) ≥ 98%. The accuracy based on the validation and study data is 100% with a 95% Confidence Interval of 97.2% to 100%.

A proportion of samples will be reported as inconclusive, as explained below. In this instance a repeat sample will be requested.

A repeat collection and test is also advised to confirm negative results in order to minimise the risk of false negatives.

How are the results interpreted?

Final patient cfDNA interpretation always takes into account the combination of results obtained for patient cfDNA (blood group allele and housekeeper) and maternal gDNA (blood group allele and housekeeper) according to the following criteria.

Result	Criteria:		
Not Detected	cfDNA:		
	 Target has ≤ 2 positive droplets 		
	 Housekeeper gene has a concentration of 7-500 copies/μL 		
	• gDNA		
	 Target has ≤ 1 positive droplets 		
	 Housekeeper gene has a concentration of 0.5-500 copies/μL 		
	Repeat collection and testing is recommended when the assay target is interpreted as 'Not Detected' and negative antigen prediction is reported.		
Detected	• cfDNA:		
	 Target has ≥ 8 positive droplets 		
	 Housekeeper gene has a concentration of 7-500 copies/μL 		
	• gDNA		
	 Target has ≤ 1 positive droplets 		
	 Housekeeper gene has a concentration of 0.5-500 copies/µL 		
Inconclusive	cfDNA:		
	 Target has 3-7 positive droplets 		
	and/or		
	 Housekeeper gene has a concentration of <7 or >500 copies/μL 		
	• gDNA		
	 Does not meet any of the criteria for gDNA listed for valid samples 		
	All cfDNA results will be considered inconclusive if the gDNA sample does not meet the validity criteria.		

How early in the pregnancy can samples be tested?

Gestational age must be at least 12 weeks.

The concentration of fetal DNA in the mother's blood increases with the progression of the pregnancy. Any sample collected before 12 weeks gestation can lead to inconclusive or false negative results and will not be tested.

Multiple Pregnancy?

We are unable to test dizygotic (fraternal) twins as it is not possible to confirm that DNA from both twins has been tested.

Clinical Indications

The Australian Red Cross Lifeblood currently offers NIPA for the following clinical indications in high risk pregnant women:

Fetal antigen	Clinical Indications		
	Alloimmunised (allo anti-D detected, regardless of titre/quantitation)		
D	2. High risk Rh(D) negative pregnant women with obstetric indications such as severe fetal maternal haemorrhage during pregnancy, or intrauterine fetal death		
D	Other unusual but rare scenarios such as allergy to the anti-D immunoglobulin		
	For 2 and 3 please discuss with a Lifeblood Transfusion Specialist prior to collection or shipping.		
K or k	Alloimmunised (anti-K or anti-k detected, regardless of titre)		
C, c, E, Fy ^a , Fy ^b	Clinically significant level of alloantibody (titre ≥ 32) or history of HDFN		

Sample Requirements

- A minimum of 3 x 6mL dedicated EDTA whole blood samples.
- Sample collection should be planned to ensure they are received by the Red Cell Reference
 Laboratory for processing within 72 hours of collection.
- Samples are only able to be processed Monday Thursday.
- Samples should be stored at 2-8°C.
- Samples must be provided with completed Red Cell Reference Laboratory request form with a minimum of 3 identifiers and include the tests requested, referring organisation's contact details and any relevant clinical information.
- The identifiers on the sample must match the request form exactly.

Samples should be sent packaged with a cold ice-brick to the QLD Red Cell Reference Laboratory, via the address below as soon as possible following collection.

QLD Red Cell Reference Laboratory Australian Red Cross Lifeblood 44 Musk Avenue (delivery via Blamey Street) Kelvin Grove, Queensland, AUSTRALIA 4059

Phone: +61 7 38389493 Fax: +61 7 38389410

Reporting

It is expected that results will be reported within 10 working days from receipt of the sample. A report containing predicted blood group phenotype of the fetus will be provided by email via Secure Send only to the referring laboratory.

Enquiries

Please direct sample collection and shipment enquiries to the Red Cell Reference Laboratory on +61 7 3838 9493.

Please direct all clinical enquiries to the Transfusion Specialist for your state. The contact details are available using this link <u>Contact | Lifeblood</u>.